

Short Communication

Advantages and disadvantages of rifampicin use in orthopedic patients to avoid *Clostridium difficile* infections

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Clostridium difficile (new taxonomy *Clostridioides difficile* [1]) is a gram-positive spore forming rod shaped bacterium being the main cause for nosocomial diarrhea. *C. difficile* infection (CDI) may include mild to severe symptoms up to fulminant colitis resulting in toxic megacolon, sepsis, multiorgan dysfunction, permanent impairment contributing to a high mortality rate. Antibiotics are thought to be the main driver for CDI, as *C. difficile* can thrive when the gut microbiome is disrupted [2]. Classically the “4 C” antibiotics clindamycin, fluoroquinolones, cephalosporins and aminopenicillins with beta-lactamase inhibitors (e.g. amoxicillin/clavulanic acid) are associated with high risk of CDI development [3]. Antibiotics with a lower risk of CDI are tetracyclins (e.g. doxycycline) and tigecyclin as well as vancomycin and daptomycin [3] CDI is treated with metronidazole, vancomycin and fidaxomicin and resistance testing is usually not necessary [4] For epidemiological purposes isolates can be differentiated by their Ribotype (RT) [5]. In recent years virulent RTs such as RT027 showed global spreading [6] leading to a higher CDI incidence and more severe courses of disease.

Virulence factors of this pathogen are the toxins A (*tcdA*) and B (*tcdB*) [7]. Some isolates inhabit additionally the binary toxin which is encoded by *cdtA* and *cdtB* [7]. Binary toxin is found preferably in epidemic isolates associated with more severe infections [7]. One of these strains is RT027 which has showed global spreading in recent years [8]. An important factor for selection and spreading of virulent strains like RT027 is their

resistance towards a variety of antibiotics, mainly macrolides and fluoroquinolones [9]. In orthopedic surgery often antibiotics are administered over long periods due to complicated soft tissue or osteoarticular infections resulting in an increased risk for CDI. In recent years an increase in CDI incidence has been noted in orthopedic patients [10] with a CDI rate of 0.19% in one major study including more than 100.000 patients [11]. Especially in patients with non-elective procedures (e.g. following trauma) CDI incidence is much more prevalent [11,12].

Rifampicin is a drug frequently used in orthopedics. The mode of action is inhibition of bacterial RNA polymerases [13] while resistance might be triggered by mutations in the *rpoB* gene in *C. difficile* isolates [14]. One major field of use are infections with biofilm formations due to staphylococci [15].

In the past it has been proposed that rifampicin might hold a protective effect towards CDI development [14,16,17]. This effect might be favored by the usually low resistance rate of *C. difficile* strains against rifampicin [18]. On the other hand other data indicate that rifampicin might favor CDI [19], especially with *C. difficile* strains harboring rifampicin resistance. In German patients suffering from osteoarticular infections rifampicin resistant RT027 isolates seemed to be a major driving factor for CDI [20]. In this study application of antibiotic stewardship which resulted in a reduction of overall antimicrobial use was able to significantly lower the CDI incidence. Similarly, in Poland, another study concluded that in an epidemiological setting with a dominance of a rifampicin resistant *C. difficile* strain (RT046) patients on tuberculosis treatment with

rifampicin are at a higher risk do develop CDI [21].

Thus, rifampicin might work as a double-edged sword: In rifampicin sensitive RTs it might protect from CDI, however, it might select rifampicin resistant *C. difficile* strains, and these are often strains with a higher virulence such as RT001, RT017, RT027 and RT176 [18]. The main goal of preventing CDI is to reduce the antibiotic use in general with a switch to drugs associated with a low risk of inducing CDI (e.g. tetracyclins). As a consequence, rifampicin should be used restrictively in orthopedic surgery and other medical fields to avoid selection of emerging resistant *C. difficile* strains and CDI development. In case of an increased CDI rate on wards with a relevant rifampicin use, ribotyping and resistance testing might be an option to identify driving factors for CDI development and to optimize antibiotic treatment.

References

1. Lawson PA, Citron DM, Tyrrell KL, Finegold SM (2016) Reclassification of *Clostridium difficile* as *Clostridioides difficile* (Hall and O'Toole 1935) *Prevot* 1938. *Anaerobe* 40: 95-99.
2. Rupnik M, Wilcox MH, Gerding DN (2009) *Clostridium difficile* infection: new developments in epidemiology and pathogenesis. *Nat Rev Microbiol* 7: 526-536.
3. Leffler DA and Lamont JT (2015) *Clostridium difficile* infection. *N Engl J Med* 372: 1539-1548.
4. Cohen SH, Gerding DN, Johnson S, Kelly CP, Loo VG, et al. (2010) Clinical practice guidelines for *Clostridium difficile* infection in adults: 2010 update by the society for healthcare epidemiology of America (SHEA) and the infectious diseases society of America (IDSA). *Infect Control Hosp Epidemiol* 31: 431-455.
5. Indra A, Huhulescu S, Schneeweis M, Hasenberger P, Kernbichler S, et al. (2008) Characterization of *Clostridium difficile* isolates using capillary gel electrophoresis-based PCR ribotyping. *J Med Microbiol* 57: 1377-1382.
6. He M, Miyajima F, Roberts P, Ellison L, Pickard DJ, et al. (2013) Emergence and global spread of epidemic healthcare-associated *Clostridium difficile*. *Nat Genet* 45: 109-113.
7. Gerding DN, Johnson S, Rupnik M, Aktories K (2014) *Clostridium difficile* binary toxin CDT: mechanism, epidemiology, and potential clinical importance. *Gut Microbes* 5: 15-27.
8. Yakob L, Riley TV, Paterson DL, Marquess J, Magalhaes RJ, et al. (2015) Mechanisms of hypervirulent *Clostridium difficile* ribotype 027 displacement of endemic strains: an epidemiological model. *Sci Rep* 5: 12666.
9. Tenover FC, Tickler IA, Persing DH (2012) Antimicrobial-resistant strains of *Clostridium difficile* from North America. *Antimicrob Agents Chemother* 56: 2929-2932.
10. Anderson PA, Bernatz J, Safdar N (2017) *Clostridium difficile* Infection: An Orthopaedic Surgeon's Guide to Epidemiology, Management, and Prevention. *J Am Acad Orthop Surg* 25: 214-223.
11. Li X, Wilson M, Nylander W, Smith T, Lynn M, et al. (2016) Analysis of Morbidity and Mortality Outcomes in Postoperative *Clostridium difficile* Infection in the Veterans Health Administration. *JAMA Surg* 151: 314-322.
12. Campbell KA, Phillips MS, Stachel A, Bosco JA, 3rd, Mehta SA (2013) Incidence and risk factors for hospital-acquired *Clostridium difficile* infection among inpatients in an orthopaedic tertiary care hospital. *J Hosp Infect* 83: 146-149.
13. Wehrli W (1983) Rifampin: mechanisms of action and resistance. *Rev Infect Dis* 3: S407-411.
14. O'Connor JR, Galang MA, Sambol SP, Hecht DW, Vedantam G, et al. (2008) Rifampin and rifaximin resistance in clinical isolates of *Clostridium difficile*. *Antimicrob Agents Chemother* 52: 2813-2817.
15. Jacqueline C and Caillon J (2014) Impact of bacterial biofilm on the treatment of prosthetic joint infections. *J Antimicrob Chemother* 1: 37-40.
16. Miller MA, Blanchette R, Spigaglia P, Barbanti F, Mastrantonio P (2011) Divergent rifamycin susceptibilities of *Clostridium difficile* strains in Canada and Italy and predictive accuracy of rifampin Etest for rifampicin resistance. *J Clin Microbiol* 49: 4319-4321.
17. Schindler M, Bernard L, Belaieff W, Gamulin A, Racloz G, et al. (2013) Epidemiology of adverse events and *Clostridium difficile*-associated diarrhea during long-term antibiotic therapy for osteoarticular infections. *J Infect* 67: 433-438.
18. Freeman J, Vernon J, Morris K, Nicholson S, Todhunter S, et al. (2015) Pan-European longitudinal surveillance of antibiotic resistance among prevalent *Clostridium difficile* ribotypes. *Clin Microbiol Infect* 21: e9-e16.
19. Choi JM, Kim HH, Park SJ, Park MI, Moon W (2011) Development of pseudomembranous colitis four months after initiation of rifampicin. *Case Rep Gastroenterol* 2011: 5.
20. Färber J, Illiger S, Berger F, Gärtner B, von Müller L, et al. (2017) Management of a cluster of *Clostridium difficile* infections among patients with osteoarticular infections. *Antimicrobial Resistance & Infection Control* 6: 22.
21. Obuch-Woszczatynski P, Dubiel G, Harmanus C, Kuijper E, Duda U, et al. (2013) Emergence of *Clostridium difficile* infection in tuberculosis patients due to a highly rifampicin-resistant PCR ribotype 046 clone in Poland. *Eur J Clin Microbiol Infect Dis* 32: 1027-1030.